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Note

Gas-liquid chromatography of methyl esters of natural penicillins

B. MEESCHAERT, P. ADRIAENS and H. EYSSEN

The Rega Institute, University of Leuven, Minderbroedersstraat 10, B-3000 Leuven (Belgium)

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During a study on the different types of penicillins produced by several mutants of *Penicillium chrysogenum*, a rapid analytical method was needed. Chromatography on water-saturated, buffered paper strips with diethyl ether as the mobile phase has been used extensively for the separation of natural penicillins in culture broths¹⁻³; however, this technique is time consuming and gives very variable results. Although a few workers have described the gas chromatographic separation of esters of semi-synthetic penicillins⁴⁻⁶, a similar method for naturally occurring penicillins is not available. In this paper we describe the separation of methyl esters of benzylpenicillin, phenoxymethylpenicillin and six penicillins with aliphatic side-chains by gas-liquid chromatography (GLC), and its application to the analysis of culture broths.

EXPERIMENTAL

Benzyl- and phenoxymethylpenicillin were obtained from R.I.T. (Genval, Belgium). Methyl-, *n*-propyl-, *n*-pentyl-, *n*-heptyl- and *n*-nonylpenicillin were prepared by condensing the appropriate acid chloride with 6-aminopenicillanic acid. The penicillins were extracted into diethyl ether from aqueous solutions at pH 2 and esterified with a slight excess of diazomethane at 0-2°; the ether solutions of the methyl esters were evaporated to dryness and the residues were dissolved in acetone. Samples of 1-3 μ l were analyzed on a Pye 104 gas chromatograph with 150-cm columns of 3% OV-1 or 3% OV-17. Temperature programmes for both columns were used; after 10 min at 150° (OV-1 column) or 180° (OV-17 column), the temperature was increased at the rate of 4°/min for 10 min and then held constant to the end of the run.

RESULTS AND DISCUSSION

Chromatograms of a synthetic mixture of the penicillin methyl esters studied are shown in Fig. 1a for the OV-1 column and in Fig. 1b for the OV-17 column. Good separations were obtained on both columns. The retention times of the aliphatic penicillins increased with the length of the side-chain; also, benzylpenicillin was eluted before phenoxymethylpenicillin. However, the elution patterns on the two columns were not identical: the separation of *n*-heptyl- and benzylpenicillin was better on the

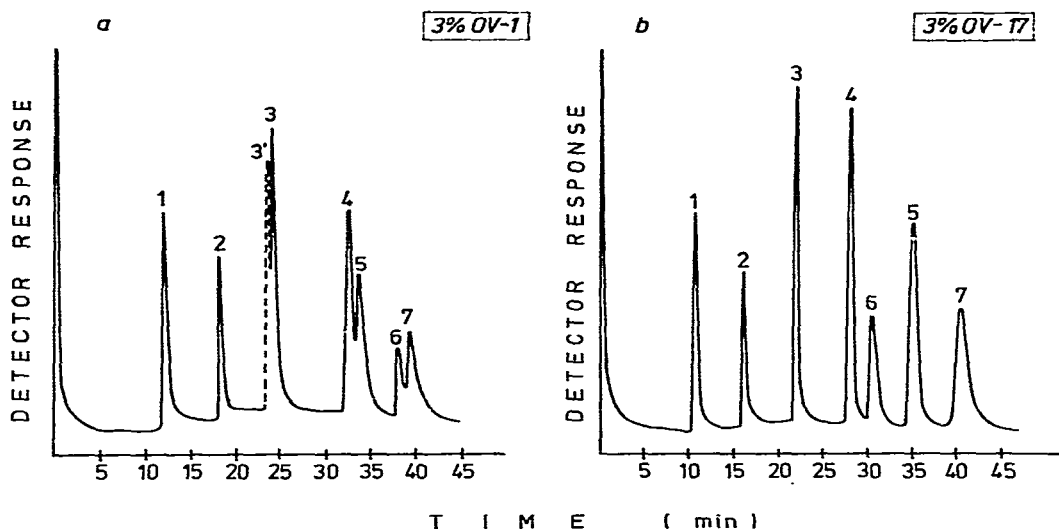


Fig. 1. Gas chromatograms of a synthetic mixture of methyl esters of naturally occurring penicillins. Coiled glass columns, 150 cm \times 4 mm I.D., packed with (a) 3% OV-1 or (b) 3% OV-17 on Gas-Chrom Q (100–120 mesh) (Applied Science Labs., State College, Pa., U.S.A.) were used. The carrier gas was nitrogen at a flow-rate of 60 ml/min; details of the temperature programme are given in the text. Peaks: 1 = methylpenicillin; 2 = *n*-propylpenicillin; 3 = *n*-pentylpenicillin; 3' = *n*-2-pentenylpenicillin; 4 = *n*-heptylpenicillin; 5 = benzylpenicillin; 6 = *n*-nonylpenicillin; 7 = phenoxy-methylpenicillin.

OV-17 column, and *n*-nonylpenicillin was eluted before benzylpenicillin on the OV-17 column and just before phoxymethylpenicillin on the OV-1 column.

The method was used to identify the different penicillins produced by several mutants of *P. chrysogenum*. Culture filtrates of *P. chrysogenum* ATCC 26818, a high-producing strain, grown in the medium of Jarvis and Johnson⁷ without added side-chain precursor, were saturated with ammonium sulphate and extracted with ethyl acetate at pH 2; the extracted penicillins were esterified with diazomethane and analysed by GLC. On the OV-17 column, *n*-pentyl- and *n*-heptylpenicillin could be detected. However, on the OV-1 column, a third penicillin was eluted just before *n*-pentylpenicillin and was identified by mass spectrometry as *n*-2-pentenylpenicillin (m/e^+ 326), which is known to occur in *Penicillium* broths². To confirm that these compounds were penicillins, a portion of the culture filtrate was first treated with penicillinase; after extraction and esterification, the penicillin methyl esters were no longer detected.

For analysis of low-yielding *Penicillium* mutants producing not more than 5 $\mu\text{g/ml}$ and grown in a rich medium⁸, the penicillinase test was very useful for the differentiation of penicillins and impurities in the extract. Ethyl acetate extracts of culture filtrates from a leu-met-cys auxotroph of *P. chrysogenum* Wis. 49-2105 were analyzed by GLC, before and after treatment with penicillinase. Although grown in the presence of phenylacetic acid, this strain produced relatively high levels of *n*-propyl-, *n*-pentyl-, *n*-2-pentenyl- and *n*-heptylpenicillin, in addition to penicillin G. In addition, an unknown penicillin was eluted between *n*-pentyl- and *n*-heptylpenicillin on the OV-1 column, but coincided with the latter on the OV-17 column. Further studies on the identification of this penicillin are in progress.

REFERENCES

- 1 R. R. Goodall and A. A. Levi, *Nature (London)*, 158 (1946) 675.
- 2 W. A. Winsten and A. H. Spark, *Science*, 106 (1947) 192.
- 3 M. L. Karnovsky and M. J. Johnson, *Anal. Chem.*, 21 (1949) 1125.
- 4 H. Vanderhaeghe, E. Evrard and M. Claesen, *Ber. XXIII Int. Kongr. Pharmaz. Wissensch., Munster, 9-14 Sept., 1963*, 1964, p. 405.
- 5 E. Evrard, M. Claesen and H. Vanderhaeghe, *Nature (London)*, 201 (1964) 1124.
- 6 C. Hishta, D. L. Mays and M. Garofalo, *Anal. Chem.*, 43 (1971) 1530.
- 7 F. G. Jarvis and M. J. Johnson, *J. Amer. Chem. Soc.*, 69 (1947) 3010.
- 8 P. A. Fawcett, J. J. Usher and E. P. Abraham, *Biochem. J.*, 151 (1975) 741.